RESEARCH LETTERS

- Krief I, Goldblatt JG, Paz A, Potasman I. Late vaccination against yellow fever of travelers visiting endemic countries. Travel Med Infect Dis. 2006;4:94

 –8. http://dx.doi.org/10.1016/j.tmaid.2005.02.001
- Gotuzzo E, Yactayo S, Córdova E. Efficacy and duration of immunity after yellow fever vaccination: systematic review on the need for a booster every 10 years. Am J Trop Med Hyg. 2013;89:434

 –44. 10.4269/ajtmh.13-0264 http://dx.doi.org/10.4269/aitmh.13-0264
- Collaborative group for studies on yellow fever vaccines. Duration of post-vaccination immunity against yellow fever in adults. Vaccine. 2014;39:4977–84. PubMed https://doi.org/ 10.1016/j.vaccine.2014.07.021

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Enterovirus A71 Infection, Thailand, 2017

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DOI: https://doi.org/10.3201/eid2407.171923

An outbreak of hand, foot and mouth disease among children in Thailand peaked in August 2017. Enterovirus A71 subgenogroup B5 caused most (33.8%, 163/482) cases. Severe disease (myocarditis and encephalitis) was observed in 1 patient. Coxsackievirus A6 was detected in 6.0% (29/482) of patients, and coxsackievirus A16 was detected in 2.7% (13/482) of patients.

Hand, foot and mouth disease (HFMD) and herpangina, caused primarily by enterovirus A, commonly affect children and result in painful blisters in the buccal cavity and on the soles of the hands and feet. In rare situations, enterovirus infection can lead to severe neurologic complications, notably aseptic meningitis, encephalitis, acute flaccid paralysis, and death in young children (1).

A nationwide outbreak of HFMD caused by coxsackievirus A6 (CV-A6) affected many children in Thailand in 2012 (2). Since that time, \approx 50,000 cases of HFMD have been reported annually to the Thailand Ministry of Public Health (3); CV-A6 and CV-A16 are the main causative agents. Most reports were based on clinical symptoms; laboratory-based confirmations are rare. Consequently, the incidence increased awareness and the need for diagnostic-based epidemiologic surveillance. As of September 25, 2017, a total of 59,071 cases of HFMD have been reported (4). We determined the prevalence and viral etiology of enterovirus infections among patients with clinical HFMD and herpangina in Thailand during 2017.

The study was approved by the Chulalongkorn University Faculty of Medicine Institutional Review Board (institutional review board no. 002/60). We tested clinical specimens from 482 children requiring hospital care for HFMD (n = 435) and herpangina (n = 47) that were submitted to King Chulalongkorn Memorial Hospital (Bangkok, Thailand) during January 1–October 30, 2017, from 12 provinces with reported HFMD outbreaks (online Technical Appendix Figure 1, https://wwwnc.cdc.gov/EID/article/24/7/17-1923-Techapp1.pdf). The study population was 273 boys and 209 girls (sex ratio M:F 1.3:1; age range 4 days to 45 years; mean \pm SD age 2.9 ± 4.0 years; median age 2.0 years).

We subjected specimens to 2 real-time reverse transcription PCRs (RT-PCRs). The first RT-PCR identifies enterovirus A71 (EV-A71), CV-A6, and CV-A16 (5). We then subjected virus-positive samples to full-length virus capsid protein 1 (VP1) gene amplification by using a conventional RT-PCR and nucleotide sequencing to identify EV-A71 subgenogroups (6). The second RT-PCR is a panenterovirus real-time RT-PCR, which also detects the glyceraldehyde-3-phosphate dehydrogenase gene (internal control). We subjected samples to a conventional RT-PCR that used CODEHOP degenerate primers to identify enterovirus serotypes other than EV-A71, CV-A6, and CV-A16 (7). We also performed molecular typing by using phylogenetic analysis and nucleotide sequence comparisons of strains. Sequences were deposited in GenBank (accession nos. MG843892-844136).

Enterovirus infections were detected throughout the year; increased frequency was observed in the rainy season (June–September) (online Technical Appendix Figure 1). Enteroviruses were identified in 67.4% (325/482) of samples. EV-A71 was most frequent (33.8%, 163/482), followed by CV-A6 (6.0%, 29/482), and CV-A16 (2.7%, 13/482). Analysis of partial VP1 sequences showed that almost all (99.4%, 162/163) of EV-A71 was subgenogroup B5: only 2 samples were subgenogroup C4 (Figure 1A). Full-length VP1 analysis also confirmed our EV-A71 subgenogroup assignment (online Technical Appendix Figure 2).

Comparison of full-length VP1 nucleotide sequences of subgenogroup B5 viruses between strains isolated in Thailand during this study and those previously isolated in Thailand showed 93.3%–100.0% identity (online Technical Appendix Table). Strains isolated in this study also had similar sequence identity with strains previously isolated (92.7%–98.6%). CV-A4 accounted for 6.6% (32/482) of isolates, followed by CV-A2 (1.5%, 7/482) and CV-A8 (1.0%, 5/482). Minor genotypes (CV-A5, CV-A10, and echovirus 9) accounted for 0.4% (2/482). One sample each was positive for CV-A9, CV-B2, echovirus 4, and echovirus 5. The remaining 13.5% (65/482) of isolates positive for panenterovirus were not typeable.

A severely ill 9-month-old boy in Nakhon Sawan Province was given a diagnosis of HFMD and required prolonged hospitalization because of sequelae caused by severe brain damage from encephalitis and myocarditis. Laboratory tests on rectal swab samples identified EV-A71 subgenogroup B5. An 18-month-old girl from Uttaradit Province who suddenly died during development of HFMD was positive for CV-A9.

Enteroviruses associated with HFMD in Thailand during 2012–2016 are usually CV-A6 or CV-A16 (5). EV-A71 has been associated with major outbreaks in China and Vietnam (8), and infections have been linked to more severe symptoms and neurologic complications in patients requiring hospitalization. In the HFMD outbreak in Thailand during 2012, deaths were primarily associated with EV-A71 subgenogroup B5 (9). The relatively high frequency of HFMD attributed to EV-A71 in this study was unexpected because our previous studies showed that <10% of enterovirus-positive samples were EV-A71 (2,9).

Our study had limitations. Because samples were obtained only from provinces with reported HFMD outbreaks, we might have missed HFMD-associated enterovirus in other regions. Limited availability of demographic and clinical data also prevented more in-depth analysis of disease severity and clinical associations. Additional deaths caused by EV-A71 infections might have been missed. Thus, without accurate and complete reporting of severe complications associated with EV-A71 infections, clinical implications from such infections could not be determined.

Effective prevention of widespread HFMD is needed. The approved EV-A71 vaccine containing subgenogroup C4 and ongoing development of B2- and B4-based vaccines can elicit cross-neutralizing antibodies against circulating EV-A71 subgenogroups, which might prove to be effective in preventing subgenogroup B5 infection, which is predominant in Thailand (10). However, molecular epidemiologic surveillance of HFMD is needed to monitor enterovirus transmission.

This study was supported by the National Science and Technology Development Agency (research chair grant P-15-5004); the Center of Excellence in Clinical Virology, Chulalongkorn University (grant GCE59-00930-005); and King

Chulalongkorn Memorial Hospital. J.P. was supported by a postdoctoral fellowship from the Ratchadaphiseksomphote Fund of Chulalongkorn University.

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References

- Solomon T, Lewthwaite P, Perera D, Cardosa MJ, McMinn P, Ooi MH. Virology, epidemiology, pathogenesis, and control of enterovirus 71. Lancet Infect Dis. 2010;10:778–90. http://dx.doi.org/10.1016/S1473-3099(10)70194-8
- Puenpa J, Chieochansin T, Linsuwanon P, Korkong S, Thongkomplew S, Vichaiwattana P, et al. Hand, foot, and mouth disease caused by coxsackievirus A6, Thailand, 2012. Emerg Infect Dis. 2013;19:641–3. http://dx.doi.org/10.3201/eid1904.121666
- Bureau of General Communicable Diseases, Ministry of Public Health, Thailand. Hand, foot, and mouth disease (HFMD) in Thailand, situation update, No. 6 (2016) [cited 2017 Nov 21]. http://thaigcd.ddc.moph.go.th/en/disease alerts/view/13
- Bureau of General Communicable Diseases, Ministry of Public Health, Thailand. Hand, foot, and mouth disease (HFMD) in Thailand, situation update, No. 6 (2017) [cited 2017 Nov 21]. http://thaigcd.ddc.moph.go.th/en/disease alerts/view/35
- Puenpa J, Suwannakarn K, Chansaenroj J, Vongpunsawad S, Poovorawan Y. Development of single-step multiplex real-time RT-PCR assays for rapid diagnosis of enterovirus 71, coxsackievirus A6, and A16 in patients with hand, foot, and mouth disease. J Virol Methods. 2017;248:92–9. http://dx.doi.org/ 10.1016/j.jviromet.2017.06.013
- Mauleekoonphairoj J, Vongpunsawad S, Puenpa J, Korkong S, Poovorawan Y. Complete genome sequence analysis of enterovirus 71 isolated from children with hand, foot, and mouth disease in Thailand, 2012–2014. Virus Genes. 2015;51:290–3. http://dx.doi.org/10.1007/s11262-015-1239-0
- Nix WA, Oberste MS, Pallansch MA. Sensitive, seminested PCR amplification of VP1 sequences for direct identification of all enterovirus serotypes from original clinical specimens. J Clin Microbiol. 2006;44:2698–704. http://dx.doi.org/10.1128/ JCM.00542-06
- Geoghegan JL, Tan V, Kühnert D, Halpin RA, Lin X, Simenauer A, et al. Phylodynamics of enterovirus A71–associated hand, foot, and mouth disease in Viet Nam. J Virol. 2015;89:8871–9. http://dx.doi.org/10.1128/JVI.00706-15
- Linsuwanon P, Puenpa J, Huang SW, Wang YF, Mauleekoonphairoj J, Wang JR, et al. Epidemiology and seroepidemiology of human enterovirus 71 among Thai populations. J Biomed Sci. 2014;21:16. http://dx.doi.org/10.1186/ 1423-0127-21-16
- Chong P, Liu CC, Chow YH, Chou AH, Klein M. Review of enterovirus 71 vaccines. Clin Infect Dis. 2015;60:797–803. http://dx.doi.org/10.1093/cid/ciu852

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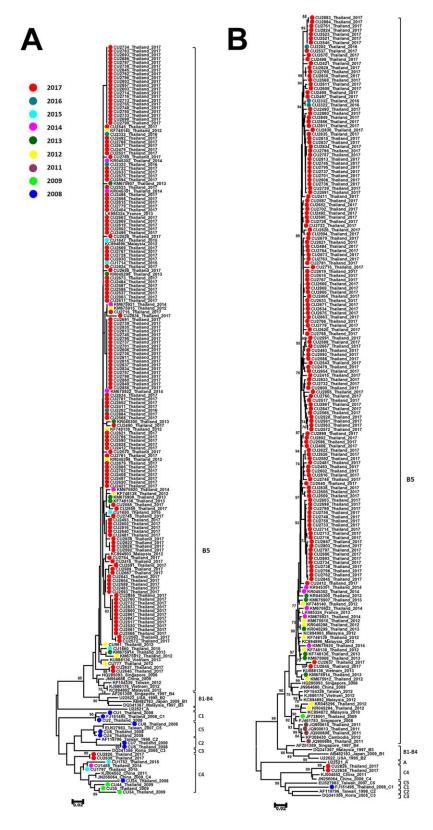
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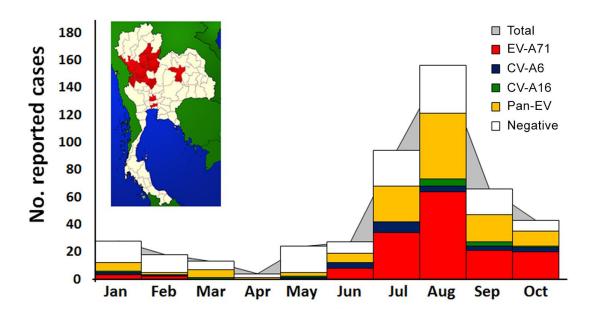
Technical Appendix Table. Comparison of nucleotide sequence identity of full-length VP1 gene of enterovirus A71 subgenogroup B5 with other virus strains*

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Sequence	CU2637	CU2412	CU2202	TH2009	TH2011	TH2013	SG2008	CN2009	MY2010	KH2012	TW2012	FR2013
CU2412	96.5											
CU2202	96.4	99.2										
TH2009	93.7	94.6	94.5									
TH2011	93.3	94.0	94.1	97.6								
TH2013	96.5	97.5	97.6	95.0	94.0							
SG2008	94.0	94.9	94.8	99.2	97.9	95.1						
CN2009	94.9	95.9	96.0	95.0	94.0	96.8	95.3					
MY2010	92.9	93.6	93.4	96.5	95.5	93.8	96.8	94.2				
KH2012	92.7	93.8	93.7	97.4	98.6	94.0	97.7	93.8	95.2			
TW2012	94.2	95.2	95.3	94.6	94.0	96.1	94.9	97.3	94.5	94.0		
FR2013	96.8	98.5	98.6	94.9	94.3	98.3	95.2	96.5	93.7	94.1	95.8	
VN2013	96.5	97.7	97.8	94.8	94.0	98.4	95.1	96.6	93.8	93.8	95.9	98.5

^{*}Values are percentage identities. Blank spaces indicate 100% identity. Strains used in phylogenetic trees and their GenBank accession numbers are TH/2009 (JF738001), TH/2011 (JQ900615), TH/2013 (KM675914), SG/2008 (FJ461783), CN/2009 (JN964686), MY/2010 (KC894872), KH/2012 (KP308430), TW/2012 (KF154339), FR/2013 (LK985324), and VN/2013 (KU888136). CN, China; FR, France; KH, Cambodia; MY, Malaysia; SG, Singapore; TH, Thailand; TW, Taiwan; VP1, virus capsid protein 1. The VP1 sequences of enterovirus A71 obtained in this study are CU2637, CU2412, and CU2202.



Technical Appendix Figure 1. Distribution of enterovirus-positive samples from case-patients with hand, foot, and mouth disease and herpangina, Thailand, January–October 2017. CV, coxsackievirus; EV, enterovirus.



Technical Appendix Figure 2. Phylogenetic analysis of A) partial and B) complete virus capsid protein 1 sequences of enterovirus A71. Virus strains were isolated from patients with hand, foot and mouth disease during 2008–2017, including those from this study, and compared with references strains from GenBank. The tree was constructed by using the neighbor-Joining method implemented in MEGA6 (https://www.megasoftware.net/). Virus strains characterized in this study are indicated by red circles. Only bootstrap resampling values ≥70% are shown. Scale bars indicate nucleotide substitutions per site.